

Determination of Leukotoxin Structural Gene (LktA) of *Mannheimia haemolytica* Isolates from Bovine Pneumonia in Japan during 2010

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ABSTRACT

This study was conducted to provide fundamental information on the capsular serotype and leukotoxin structural gene (lktA) genotypes of *Mannheimia haemolytica* field isolates from cattle with pneumonic pasteurellosis in Japan during 2010. Of the 54 isolates serotyped, 29 (53.7%) were serotype A1, 10 (18.5%) were serotype A2, 8 (14.8%) were serotype A6, and 7 (13.0%)

were untypable via slide agglutination testing. Sequencing analysis revealed that the lktA1.1-type leukotoxin was exclusively associated with serotypes A1 and A6. In contrast, lktA2.1- and 8.1-type leukotoxins were associated with both serotype A2 and untypable isolates.

This study revealed that cattle infected with *M. haemolytica* in Japan were exposed to at least three types of leukotoxins. However, further comprehensive studies are required to investigate allelic variants of

Table 1. Characterization of *M. haemolytica* used in this study. ^a

BAC plate ^b	Phenotype					PCR
Growth	L-Arabinose	Sorbitol	Aesculin	Catarase	Oxidase	<i>lkt</i>
+	-	+	-	+	+	+

^a Results are summarized based on the 54 isolates from pneumonic pasteurellosis in Japan in 2010.

^b Blood agar plates containing 15 ug/mL bacitracin.

lktA in *M. haemolytica* prevailed in Japan.

INTRODUCTION

Mannheimia haemolytica is a complicating agent in bovine respiratory disease, and is recognized as an important pathogen in feedlot cattle (Rice et al, 2007). Based on capsular antigens or lipopolysaccharide complexes, there are 17 serotypes and numerous untypable strains of *M. haemolytica* (Angen et al, 1999). Bovine pasteurellosis, primarily due to serotype A1 commonly manifests as severe fibrinous pleuropneumonia (Lillie, 1974).

M. haemolytica leukotoxin is specifically virulent for ruminant lymphoid cells, and is a key factor in the pathogenesis of pneumonic pasteurellosis. Significant antibody responses to leukotoxins correlate with resistance to experimental *M. haemolytica* challenge (Clinkenbeard et al, 1989; Highlander et al, 1989). However, the cytotoxic activity of *M. haemolytica* strains depends on the diversity of the leukotoxin structural gene (*lktA*), which has a complex evolutionary history and at least eight major allelic variants (Davies et al, 2001).

The identification of prevalent serotypes and allelic variants of *lktA* provides useful information for vaccine development. However, little is known about the distribution of *lktA* allelic variants in Japan. Thus, this study will investigate the prevalence of *M. haemolytica* serotypes and allelic variants of *lktA* throughout Japan.

MATERIALS AND METHODS

M. haemolytica Field Isolates

total of 54 *M. haemolytica* isolates were obtained from different cattle with pneumonic pasteurellosis in 16 prefectures in Japan during 2010. The isolates were analyzed for catalase and oxidase activities, and positive samples were then identified via multiplex PCR (Alexander et al, 2008).

Serotyping of *M. haemolytica* Isolates from Bovine Pneumonia

Antisera were prepared in rabbits by intravenous inoculations of formalinized whole cell suspensions of reference strains (Biberstein, 1978; Fodor et al, 1988) of *M. haemolytica*, provided by Dr. G.H. Frank, National Animal Disease Center, Ames, Iowa, USA. Briefly, two New Zealand rabbits were injected biweekly with an antigen of each serotype reference strain. Antisera were used to serotype isolates using a slide agglutination test (Frank and Wessman, 1978). However, antiserum against the newly described serotype A17 was not included in the test.

lktA Genotyping of *M. haemolytica* Isolates from Bovine Pneumonia

PCR amplification and DNA sequence analysis were conducted based on the method reported by Davies et al (2001). Namely, the *lktA* gene was amplified from the chromosomal DNA with the 5' primer

Table 2. Frequency of *M. haemolytica* isolates in Japan during 2010

Serotype				Total number of isolates
A1	A2	A6	UT ^a	
Number of isolates (%) ^b				54
29 (53.7)	10 (18.5)	8 (14.8)	7 (13.0)	

^a Untypable

^b Percentage of each serotype among the total isolates

Table 3. *LktA*-types of *M. haemolytica* isolates in Japan (*n*=32).

Serotype	A1		A2		A6		UT ^a	
	A1.1	A2.1	A8.1	A1.1	A2.1	A8.1		
Number ^b	14	2	4	8	1	3		

^a Untypable

^b Number of tested isolates

lktA9 (5'-TCA AGA AGA GCT GGC AAC-3') and the 3' primer *lktA7* (5'-AGT GAG GGC AAC TAA ACC-3'). PCRs were carried out in a thermal cycler (TAKARA, Tokyo Japan) using the following amplification parameters: denaturation at 94°C for 45 s, annealing at 62°C for 45 s, and extension at 72°C for 2 min. Thirty cycles were performed, and a final extension step of 72°C for 10 min was used. Production of a PCR amplicon of the expected size (~3 kbp) was confirmed by agarose gel electrophoresis, and the DNA was purified with a QIA-quick PCR purification kit (Qiagen, Tokyo, Japan). Dye terminator cycle sequencing was performed with the Big Dye Terminator Cycle sequencing kit (Applied Biosystems, Carlsbad, CA, USA), and sequence reactions were performed and read by an automated DNA sequencer (ABI PRISM3130, Applied Biosystems). Both strands of the *lktA* gene were sequenced using seven internal primer pairs, designed as sequence data became available. Nucleotide sequence data were analyzed and edited with the GENETYX (version 9) program.

Nucleotide Sequence Accession Numbers

The GenBank accession numbers used in this study are *lktA1.1* - AF314503, *lktA2.1* - AF314511, and *lktA8.1* - AF314515.

RESULTS

Frequency of *M. haemolytica* Serotype in Japan During 2010

All 54 field isolates from cattle with pneumonic pasteurellosis in 2010 were phenotypically and genetically identified as *M. haemolytica* (Table 1). Serotyping results are shown in Table 2. Of the 54 isolates serotyped, 29 (53.7 %) were serotype A1, 10 (18.5 %) were serotype A2, 8 (14.8 %) were

serotype A6, and 7 (13.0 %) isolates were untypable.

LktA-types of *M. haemolytica* Isolates in Japan During 2010

Of the 54 isolates, 32 (14 serotype

A1, 6 serotype A2, 8 serotype A6, 4 untypable isolates) were selected from different prefectures in order to increase geographic diversity. *lktA* allelic variants were analyzed by DNA sequencing (Table 3). Serotypes A1 and A6 all contained *lktA1.1*, and serotype A2 and untypable isolates contained either *lktA2.1* or 8.1; 22 (68.8 %), 3 (9.4 %) and 7 (21.9 %) isolates produced *lktA1.1*-, 2.1-, and 8.1-type leukotoxins, respectively.

Alignment Comparison of Putative Amino Acid Sequences for *lktA*

Putative amino acid sequences of *lktA* were aligned to previously reported sequences (data not shown). These putative amino acid sequences had 100% identity to the reference sequences previously reported by Davies et al [6].

DISCUSSION

For many bacterial pathogens, identifications of serotype and virulence factor are useful in understanding outbreak epidemiology, monitoring of infected cattle, and preparing vaccines for disease control.

In accordance with serological survey done by Katsuda et al (2012), the majority (87%) of isolates were represented by only three serotypes (A1, A2 and A6). Serotypes A1 and A6 only used *lktA1.1*, while serotype A2 and untypable isolates used *lktA8.1* or A2.1. Davies et al also reported that *lktA1.1* was distributed among serotype A1 and A6 (Davies and Baillie, 2003). Though the *lktA8.1*-type leukotoxin is well known to be an ovine isolate (Davies, 1997), they appeared to be isolated from bovine with pneumonic pasteurellosis in Japan.

Strain selection for vaccine development is pivotal for disease protection. In

Japan, two commercial vaccines have been developed; both are from serotype A1 and have prevented disease in cattle, since the prevalence of *M. haemolytica* in Japan has remained relatively constant.

CONCLUSION

However, continuous determination of serotype and *lktA* allelic variants of field isolates would aid in determination of effective control measures against *M. haemolytica*. As far as we know, this is the first report on information regarding capsular serotypes and the allelic variations of *lktA* in *M. haemolytica* recently prevailed in Japan.

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REFERENCES

1. Rice JA, Carrasco-Medina L, Hodgins DC, Shewen PE: *Mannheimia haemolytica* and bovine respiratory disease. *Anim Health Res Rev* 2007, 8: 117-128.
2. Angen O, Mutters R, Caugant DA, Olsen JE, Bisgaard M: Taxonomic relationships of the *Pasteurella haemolytica* complex as evaluated by DNA-DNA hybridizations and 16S rRNA sequencing with proposal of *Mannheimia haemolytica* gen. nov., comb. nov., *Mannheimia granulomatis* comb. nov., *Mannheimia glucosida* sp. nov., *Mannheimia ruminalis* sp. nov. and *Mannheimia varigena* sp. nov. *Int J Syst Bacteriol* 1999, 49: 67-86.
3. Lillie LE: The bovine respiratory disease complex. *Can Vet J* 1974, 15: 233-242.
4. Clinkenbeard KD, Mosier DA, Timko AL, Confer AW: Effects of *Pasteurella haemolytica* leukotoxin on cultured bovine lymphoma cells. *Am J Vet Res* 1989, 50: 271-275.
5. Highlander SK, Chidambaram M, Engler MJ, Weinstock GM: DNA sequence of the *Pasteurella haemolytica* leukotoxin gene cluster. *DNA Cell Biol* 1989, 8: 15-28.
6. Davies RL, Whittam TS, Selander RK: Sequence diversity and molecular evolution of the leukotoxin (*lktA*) gene in bovine and ovine strains of *Mannheimia* (*Pasteurella*) *haemolytica*. *J Bacteriol* 2001, 183: 1394-1404.
7. Alexander TW, Cook SR, Yanke LJ, Booker CW, Morley PS, Read RR, Gow SP, McAllister TA: A multiplex polymerase chain reaction assay for the identification of *Mannheimia haemolytica*, *Mannheimia glucosida* and *Mannheimia ruminalis*. *Vet Microbiol* 2008, 130: 165-175.
8. Biberstein EL: Biotyping and serotyping of *Pasteurella haemolytica* In: T. Bergam, J.R. Norris (Eds.), *Methods in Microbiology*, vol. 10 Academic Press, London, 1978: 253-269.
9. Fodor L, Varga J, Hajtos J, Donachie W, Gilmour NJ: Characterisation of a new serotype of *P. haemolytica* isolated in Hungary. *Res Vet Sci* 1988, 44: 399.
10. Frank GH, Wessman GE: Rapid plate agglutination procedure for serotyping *Pasteurella haemolytica*. *J Clin Microbiol* 1978, 7: 142-145.
11. Katsuda K, Kohmoto M, Mikami O. Relationship between serotype and the antimicrobial susceptibility of *Mannheimia haemolytica* isolates collected between 1991 and 2010. *Res Vet Sci*. 2013 Apr;94(2):205-8. doi: 10.1016/j.rvsc.2012.09.015. Epub 2012 Oct 11.
12. Davies RL, Baillie S: Cytotoxic activity of *Mannheimia haemolytica* strains in relation to diversity of the leukotoxin structural gene *lktA*. *Vet Microbiol* 2003, 92: 263-79.
13. Davies RL, Arkinsaw S, Selander RK: Evolutionary genetics of *Pasteurella haemolytica* isolates recovered from cattle and sheep. *Infect Immun* 1997, 65: 3585-3593.